# DAISEP SPIN XS
## PROTOCOL

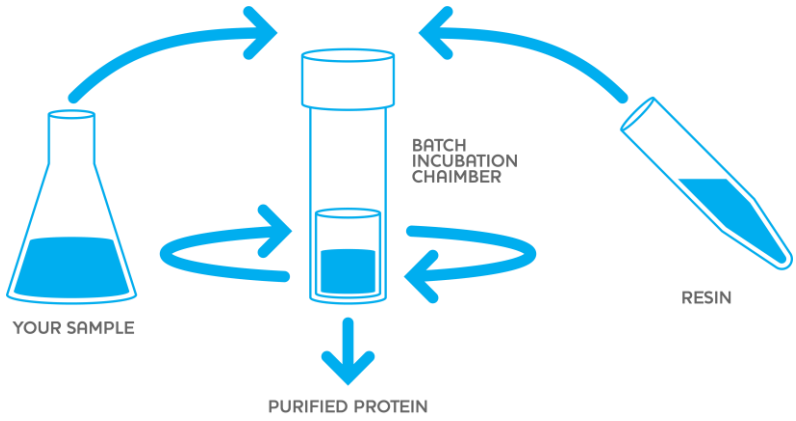
## MATERIALS SUPPLIED IN THE KIT:

- DAISEP Spin XS column (0.6 mL capacity in a swing bucket rotor)
  40 units
- 2.2 mL centrifuge tubes
  80 units

## ADDITIONAL MATERIALS REQUIRED:

- 0.2 µm clarification device
- Microfuge with a fixed angle rotor capable of handling 2.2 mL centrifuge tubes (diameter 11 mm)
- Buffer
- Quartz cuvettes for UV absorbance measurements
- UV/VIS spectrophotometer

## PROTEIN PURIFICATION PROTOCOL:

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## RECOMMENDED PROTOCOL

The following spin speeds and times are appropriate for a 100 µl resin bed volume. Spin times for each of the following steps may increase with larger bed volumes.

## PRE-EQUILIBRATION

1. Pipette the appropriate resin slurry into the batch incubation chamber of the spin column barrel. Wash the resin at 12-14,000 x g for 20 sec. This step is critical to ensure that storage solution is removed from the resin.

NOTE: Ethanol does interfere with sealing properties of the Self Seal™ membrane technology.

2. Pre-equilibrate the DAISEP Spin column with 0.6 mL equilibration buffer by centrifuging the spin column at 12-14,000 x g for 20 sec. It is **critical** that you repeat this step <u>one more time</u> with a further 0.6 mL fresh equilibration buffer.

NOTE: If using one spin column, ensure that the spin column is counterbalanced with a unit of equal weight (adjusted with distilled water).

## CLARIFICATION OF SAMPLE

3. Pre-filter the sample through a single 0.2 µm filter (e.g. syringe filter).

NOTE: It is critical that the sample is filtered through a final 0.2 µm syringe filter **immediately** before loading it on the spin column. Optimal performance of these devices will depend on these instructions being rigorously followed.

## SAMPLE LOADING

4. Load the required volume of filtered sample to reach the appropriate resin/sample mix ratio (maximum sample volume is 0.6 mL). Close the lid and mix the sample and the resin (invert 2-3 times). Place the column on a standard tube roller and mix during appropriate contact times.

5. After batch incubation, centrifuge the column at 12-14000 x g for 20 sec and collect the flowthrough fraction.

NOTE: If using one spin column, ensure that the spin column is counterbalanced with a unit of equal weight (adjusted with distilled water).

## WASHING STEPS

6. If you need to perform washing, transfer the spin column to fresh centrifuge tube and load wash buffer. Close the lid, mix, centrifuge and collect the flowthrough fraction.